

Integrated control of *Fusarium* damping-off in conifer seedlings

Integrierte Bekämpfung der durch *Fusarium* verursachten Umfallkrankheit an Kiefern-Keimpflanzen

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Summary

Interactions among three species of ectomycorrhizal fungi (*Hebeloma longicaudum*, *Laccaria bicolor* and *Paxillus involutus*), a litter decomposing fungus (*Clitocybe clavipes*), two species of root pathogen (*Fusarium moniliforme* and *F. oxysporum*), and a fungicide (oxine benzoate) were studied. In dual culture *C. clavipes* and *P. involutus* reduced growth of *F. moniliforme* and *F. oxysporum*. Culture extracts of *C. clavipes*, *L. bicolor*, and *P. involutus* reduced mycelial growth and spore germination of *F. moniliforme* and *F. oxysporum*. *Hebeloma longicaudum* showed no inhibitory effect against *F. moniliforme* and *F. oxysporum*. Two compounds, o-coumaric acid (isolated from *P. involutus*) and p-hydroxybenzoic acid (isolated from *L. bicolor*) reduced growth and spore germination of *F. moniliforme* and *F. oxysporum* up to 1000 µg/ml. Oxine benzoate at recommended dose reduced root rot severity and increased survival of lodgepole pine and white spruce seedlings against *F. moniliforme* and *F. oxysporum*. *Clitocybe clavipes*, *L. bicolor*, and *P. involutus* also improved seedling survival and reduced root rot severity when inoculated onto the seedlings alone or in combination with recommended and reduced rates of oxine benzoate.

Key words: Mycorrhizal fungi; biological control; integrated control; damping-off; conifer seedlings.

Zusammenfassung

Die Interaktionen zwischen drei Arten von Ektomykorrhiza-Pilzen (*Hebeloma longicaudum*, *Laccaria bicolor* und *Paxillus involutus*), einem streuzersetzenden Pilz (*Clitocybe clavipes*), zwei Arten von Wurzelpathogenen (*Fusarium moniliforme* und *F. oxysporum*) und einem Fungizid (Oxinbenzoat) wurden untersucht. In Doppelkulturen reduzierten *C. clavipes* und *P. involutus* das Wachstum von *F. moniliforme* und *F. oxysporum*. Kulturrextrakte von *C. clavipes*, *L. bicolor* und *P. involutus* verminderten das Myceliumwachstum und die Sporenkeimung von *F. moniliforme* und *F. oxysporum*. *Hebeloma longicaudum* zeigte keine hemmende Wirkung gegen die beiden *Fusarium*-Arten auf. Zwei Verbindungen, nämlich o-Coumarinsäure (isoliert von *P. involutus*) und p-Hydroxybenzoësäure (isoliert von *L. bicolor*) reduzierten Konzentrationen von 100 bis 1000 µg/ml das Wachstum und die Sporenkeimung von *F. moniliforme* und *F. oxysporum*. Oxinbenzoat verminderte in den empfohlenen Konzentrationen die Befallsrate der Wurzelfäule und erhöhte das Überleben der Keimpflanzen von Drehkiefern und Schimmelpilzen. *Clitocybe clavipes*, *L. bicolor* und *P. involutus* verbesserten ebenfalls das Überleben der Keimpflanzen, wenn sie einzeln mit dem empfohlenen oder reduzierten Konzentrationen von Oxinbenzoat inkuliert wurden.

Stichwörter: Mykorrhizapilze; biologische Bekämpfung; integrierte Bekämpfung; Umfallkrankheit Koniferen-Keimpflanzen

1 Introduction

In North American conifer nurseries, species of *Fusarium* are responsible for causing damping-off seedlings and result in considerable losses (BLOOMBERG 1973; SUTHERLAND et al. 1989; HIRATSUKA et al. 1995; PAIGE et al. 1995). Several fungicides are used to control the damping-off. Currently, these fungicides have become less effective due to the development of pathogen resistance (OGAWA et al. 1981; ZHENG 1982). It has been suggested by several authors that biological control, using fungi or other micro-organisms as either an alternative or supplement to fungicides, may be a viable alternative for controlling damping-off (BAKER and COOK 1982; MARX 1972, 1973; PETERSON et al. 1984; BAILEY 1987; DUCHESNE et al. 1989). Several species of ectomycorrhizal fungi are known to protect or partially protect conifer seedlings under laboratory conditions (DUCHESNE et al. 1987, 1988a,b; CHAKRABORTY et al. 1990, 1991; CHAKRABORTY and HWANG 1991; HWANG et al. 1995). However, the relative effectiveness of ectomycorrhizal fungi in disease suppression varies with mycorrhizal species or isolate host species, and soil conditions (SAMPAGNI et al. 1985).

Hebeloma longicaudum Pers.: Fr. Kummer, *Laccaria bicolor* (R. Mre.) Orton, and *Paxillus involutus* (Batsch) Fr. are the three most common species of ectomycorrhizal fungi in the prairie province of Canada (NAGASAWA, unpublised report). They can form abundant ectomycorrhizae with container-grown conifer seedlings (MOLINA 1980, 1982; SHAW et al. 1982; BOYLE et al. 1987; BOYLE and HELLENBRAND 1991) and have great potential for wide-scale nursery inoculations. *Clitocybe clavipes* (Fr.) Kummer is a saprophytic fungus that occurs both in mixed woods and coniferous forests (ARNDT 1979) and is antagonistic to certain species of root pathogens. All these fungi also occur in diverse soil habitats throughout the temperate parts of the world and can be easily isolated from sporocarps.

The objectives of this study were to investigate the protective effect of *C. clavipes*, *H. longicaudum*, *L. bicolor*, *P. involutus*, and a fungicide oxine benzoate (8-hydroxyquinoline benzoate) on lodgepole pine (*Pinus contorta* Dougl. var. *latifolia* Engelm.) and white spruce (*Picea glauca* [Moench] Voss) seedlings against *Fusarium moniliforme* Sheldon (= *Fusarium verticillioides* (Sacc.) Nirenberg) and *F. oxysporum* Schlecht alone or in combination.

2 Materials and methods

2.1 In vitro antagonism of damping-off fungi and test fungi

Antagonism of *C. clavipes* (UAMH 5580), *H. longicaudum* (UAMH 9317), *L. bicolor* (NOF 2290), *P. involutus* (NOF 2340) against two species of root pathogens, *F. moniliforme* (BCRI-F45) and *F. oxysporum* (BCRI-F30), was studied on Modified Melin Norkrans (MMN) medium (MARX 1969) in 90-mm Petri plates. *Clitocybe clavipes*, *H. longicaudum*, *L. bicolor* and *P. involutus* were inoculated separately, placing 5-mm agar plugs at the margin of the plates. They were allowed to grow at 20 °C in the dark. After 7 days, 5-mm mycelial plugs of *F. moniliforme* and *F. oxysporum* were placed on the plate opposite to growing colonies of the test fungi. The antagonism was observed after 5 days and photographed.

2.2 Effect of culture extract of test fungi on mycelial growth and spore germination of *Fusarium* spp.

Clitocybe clavipes, *H. longicaudum*, *L. laccata* and *P. involutus* were grown in 250 ml liquid MMN medium at 20 °C in the dark on a shaker. After 24 days, culture filtrate was collected by filtration through Whatman No. 1 filter paper. The filtrate (200 ml) was concentrated under vacuo to 5 ml and then extracted with ethyl acetate. Ethyl acetate extract was concentrated under vacuo to dryness. Five concentrations (1, 10, 100, and 1000 µg/ml) of culture extracts were made using sterile distilled water and were stored at 2 °C for bioassay. p-hydroxybenzoic acid and o-coumaric acid were isolated from culture filtrate of *L. bicolor* and *P. involutus*, respectively. The culture filtrate (5 l) was concentrated

watered with 300 ml of aqueous solution (100 %, 50 %, or 25 % of recommended dose) every 15 d. No fertilizer was used. Seedlings were harvested and evaluated 8 weeks later. Seedling survival, root severity, mycorrhizal short roots, and total dry weight of each seedling were determined. Root severity was based on a scale: 0 = no root rot symptoms, 4 = severely necrotic root system.

2.5 Data analysis

Data from all the experiments were subjected to analysis of variance. The individual means were compared using Scheffe's test for multiple comparison using SAS software (SAS INSTITUTE INC. 1990).

3 Results

3.1 Interactions of *F. moniliforme* and *F. oxysporum* and test fungi

In dual culture, *P. involutus* and *C. clavipes* reduced the growth of *F. moniliforme* and *F. oxysporum* (Fig. 1). Both *F. moniliforme* and *F. oxysporum* had mutual inhibition against *L. bicolor*, i. e., growth of *F. moniliforme* and *F. oxysporum* and *L. bicolor* stopped when they came in contact with each other. Both *F. moniliforme* and *F. oxysporum* grew over *H. longicaudum*.

3.2 Effect of culture extract of test fungi on *F. moniliforme* and *F. oxysporum*

Mycelial growth of *F. moniliforme* and *F. oxysporum* was significantly reduced when treated with culture extracts of *C. clavipes*, *L. bicolor*, and *P. involutus* and o-coumaric acid at concentrations of 5 to 50 µg/ml (Table 1). For p-hydroxybenzoic acid, mycelial growth of *F. moniliforme* and *F. oxysporum* was reduced at 50 µg/ml. The culture extract of *H. longicaudum* had no inhibitory effect against *F. moniliforme* and *F. oxysporum* (Table 1).

Spore germination of *F. moniliforme* and *F. oxysporum* was significantly reduced when treated with culture extracts of *C. clavipes*, *L. bicolor*, and *P. involutus* and o-coumaric acid and p-hydroxybenzoic acid at 100 to 1000 µg/ml (Table 2). The culture extract of *H. longicaudum* had no inhibitory effect on spore germination of *F. moniliforme* and *F. oxysporum* (Table 2).

3.3 Effect of oxine benzoate on the *In vitro* growth of test fungi and *F. moniliforme* and *F. oxysporum*

Oxine benzoate reduced *In vitro* mycelial growth of *F. moniliforme* and *F. oxysporum* at 1.7 µg/ml. The growth of *C. clavipes*, *H. longicaudum*, *L. bicolor*, and *P. involutus* was reduced at 1700 µg/ml (Table 3).

3.4 Effect of oxine benzoate on seedling mortality, growth, disease development, and ectomycorrhiza formation of lodgepole pine and white spruce

When inoculated with *F. moniliforme* and *F. oxysporum*, seedling survival of lodgepole pine was 40 and 44 %, respectively (Table 4). For white spruce, seedling survival was 33 % and 35 % when inoculated with *F. moniliforme* and *F. oxysporum* (Table 5). When co-inoculated with *C. clavipes*, *L. bicolor*, *P. involutus* and oxine benzoate at recommended dose and 50 % of the recommended dose, seedling survival was increased and root rot severity was reduced (Tables 4 and 5). However, oxine benzoate at one-quarter of the recommended dose (25 %) had no effect on seedling survival and root rot was similar to that of seedlings inoculated with the pathogens alone. Highest seedling survival and low root rot of lodgepole pine and white spruce were observed when seedlings were treated with 1/2 recommended dose of oxine benzoate alone or in combination with *C. clavipes*, *L. bicolor*, or *P. involutus* (Tables 4 and 5). Mycorrhizal colonization was reduced when seedlings were inoculated with *F. moniliforme* and *F. oxysporum* at recommended rates and half rates of oxine benzoate. However, mycorrhizal colonization in lodgepole pine was not decreased when treated with 25 % of the recommended rates of oxine benzoate (Table 4). Total dry weight of seedlings was reduced when treated with *F. moniliforme* and *F. oxysporum* alone. The growth of seedlings was improved when treated with recommended rates of oxine benzoate.

Table 1. Effect of culture extracts of test fungi and their metabolites on the *in vitro* mycelial growth of *Fusarium moniliforme* and *F. oxysporum*Tab. 1. Wirkung von Kulturextrakten von den Testpilzen und ihren Metaboliten auf das *in vitro*-Wachstum von *F. moniliforme* und *F. oxysporum*

Concentration ($\mu\text{g/ml}$)	Mycelial dry wt. (mg)					
	Pi	Lb	Cc	Hl	OCA	PHB
<i>F. moniliforme:</i>						
0	185 ^a	182 ^a	184 ^a	185 ^a	182 ^a	18 ^a
0.05	186 ^a	182 ^a	183 ^a	185 ^a	183 ^a	18 ^a
0.5	186 ^a	182 ^a	182 ^a	186 ^a	182 ^a	18 ^a
5	173 ^b	182 ^a	162 ^b	184 ^a	169 ^b	18 ^a
50	163 ^c	164 ^b	148 ^c	185 ^a	161 ^c	16 ^c
<i>F. oxysporum:</i>						
0	186 ^a	180 ^a	181 ^a	185 ^a	181 ^a	18 ^a
0.05	185 ^a	183 ^a	182 ^a	185 ^a	181 ^a	18 ^a
0.5	185 ^a	182 ^a	182 ^a	185 ^a	181 ^a	18 ^a
5	172 ^b	175 ^b	174 ^b	170 ^b	170 ^b	18 ^a
50	164 ^c	166 ^c	165 ^c	185 ^a	161 ^c	16 ^c

Means within a column for a particular pathogen followed by the same letter are not significantly ($P \leq 0.05$) different from each other by Scheffé's test.
Pi = *Paxillus involutus*, Lb = *Laccaria bicolor*, Cc = *Clitocybe clavipes*, Hl = *Hebeloma longicaudum*, OCA = o-coumaric acid, PHBA = p-hydroxybenzoic acid.

Table 2. Effect of culture extracts of test fungi and their metabolites on spore germination of *Fusarium moniliforme* and *F. oxysporum*Tab. 2. Wirkung der Kulturextrakte von den Testpilzen und ihren Metaboliten auf die Sporenkeimung von *F. moniliforme* und *F. oxysporum*

Concentration ($\mu\text{g/ml}$)	Spore germination (%)					
	Pi	Lb	Cc	Hl	OCA	PHB
<i>F. moniliforme:</i>						
0	96 ^a	90 ^a	91 ^a	95 ^a	95 ^a	94
1	95 ^a	90 ^a	90 ^a	95 ^a	96 ^a	95
10	95 ^a	90 ^a	90 ^a	95 ^a	95 ^a	95
100	75 ^b	90 ^a	83 ^b	96 ^a	70 ^b	81
1000	49 ^c	66 ^b	73 ^c	95 ^a	44 ^c	63
<i>F. oxysporum:</i>						
0	92 ^a	90 ^a	91 ^a	91 ^a	90 ^a	90
1	92 ^a	90 ^a	91 ^a	92 ^a	91 ^a	91
10	92 ^a	91 ^a	91 ^a	91 ^a	91 ^a	90
100	75 ^b	84 ^b	84 ^b	91 ^a	70 ^b	83
1000	39 ^c	74 ^c	75 ^c	91 ^a	35 ^c	72

Means within a column for a particular pathogen followed by the same letter are not significantly ($P \leq 0.05$) different from each other by Scheffé's test.
Pi = *Paxillus involutus*, Lb = *Laccaria bicolor*, Cc = *Clitocybe clavipes*, Hl = *Hebeloma longicaudum*, OCA = o-coumaric acid, PHBA = p-hydroxybenzoic acid.

use of ectomycorrhizal fungi in the biological control of root pathogenic fungi has been discussed by ZAK (1964), MARX (1972, 1973), DUCHESNE et al. (1989), CHAKRAVARTY and UNESTAM (1987a, and CHAKRAVARTY et al. (1991). Several authors have proposed a mechanism of disease suppression by ectomycorrhizal fungi. MARX (1969) and MARX and DAVEY (1969) suggested that the introduction of disease resistance by ectomycorrhizal fungi is associated with either the formation of a physical barrier

Table 4. Interactions of test fungi and *Fusarium moniliforme* and *F. oxysporum* on seedling survival, growth, root rot severity, and ectomycorrhizal development on lodgepole pine seedlings
 Tab. 4. Interaktionen der Testpilze und *F. moniliforme* und *F. oxysporum* auf das Überleben der Keimpflanze, auf das Wachstum, auf die Schwere der Wurzelfäule und auf die Ektomykorrhiza-Entwicklung von Keimlingen der Drehkiefer

Treatment	Seedling Survival (%)	Root Rot Index	Mycorrhizal short roots (%)	Total dry wt. (g)
Control	100 ± 0	0 ± 0	0 ± 0	851 ± 5.
Pi	100 ± 0	0 ± 0	90 ± 1.5	840 ± 4.
Lb	100 ± 0	0 ± 0	85 ± 2.0	865 ± 2.
Cc	100 ± 0	0 ± 0	0 ± 0	844 ± 4.
Fm	40 ± 2.0	2.6 ± 0.7	0 ± 0	114 ± 3.
Fo	44 ± 2.0	3.4 ± 0.7	0 ± 0	212 ± 2.
Pi + Fm	69 ± 2.3	2.0 ± 0.8	61 ± 1.2	454 ± 5.
Lb + Fm	70 ± 2.3	2.6 ± 0.8	63 ± 1.5	390 ± 3.
Cc + Fm	68 ± 1.8	1.8 ± 0.6	0 ± 0	410 ± 2.
Pi + Fo	72 ± 1.8	1.6 ± 0.7	50 ± 1.3	472 ± 3.
Lb + Fo	70 ± 1.6	1.8 ± 0.9	54 ± 1.7	453 ± 5.
Cc + Fo	77 ± 1.8	1.4 ± 0.7	0 ± 0	421 ± 5.
Fm + Ob (100 %)	95 ± 1.3	1.0 ± 0.8	0 ± 0	549 ± 5.
Fm + Ob (50 %)	63 ± 1.5	2.5 ± 1.0	0 ± 0	512 ± 3.
Fm + Ob (25 %)	35 ± 1.8	3.0 ± 1.0	0 ± 0	233 ± 3.
Fo + Ob (100 %)	98 ± 1.1	0.5 ± 0.8	0 ± 0	535 ± 4.
Fo + Ob (50 %)	60 ± 1.3	2.5 ± 1.0	0 ± 0	499 ± 2.
Fo + Ob (25 %)	38 ± 1.6	3.0 ± 1.0	0 ± 0	250 ± 3.
Fm + Pi + Ob (100 %)	95 ± 1.3	0.5 ± 0.7	70 ± 2.0	720 ± 4.
Fm + Lb + Ob (100 %)	96 ± 1.0	0.1 ± 0.8	64 ± 2.2	710 ± 2.
Fm + Cc + Ob (100 %)	93 ± 1.3	0.5 ± 0.8	0 ± 0	700 ± 2.
Fm + Pi + Ob (50 %)	77 ± 1.6	1.5 ± 0.8	85 ± 1.7	610 ± 3.
Fm + Lb + Ob (50 %)	81 ± 1.1	1.5 ± 1.0	77 ± 1.8	593 ± 2.
Fm + Cc + Ob (50 %)	83 ± 1.5	0.5 ± 0.8	0 ± 0	630 ± 3.
Fm + Pi + Ob (25 %)	45 ± 1.7	2.0 ± 0.9	90 ± 1.7	550 ± 3.
Fm + Lb + Ob (25 %)	50 ± 1.5	2.5 ± 1.0	85 ± 1.7	534 ± 3.
Fm + Cc + Ob (25 %)	48 ± 1.7	2.0 ± 1.0	0 ± 0	570 ± 3.
Fo + Pi + Ob (100 %)	97 ± 1.2	0.5 ± 0.8	66 ± 1.9	709 ± 2.
Fo + Lb + Ob (100 %)	98 ± 0.9	0.3 ± 0.6	65 ± 2.0	697 ± 1.
Fo + Cc + Ob (100 %)	95 ± 1.8	0.2 ± 0.6	0 ± 0	700 ± 1.
Fo + Pi + Ob (50 %)	60 ± 1.2	1.0 ± 1.0	75 ± 2.0	591 ± 1.
Fo + Lb + Ob (50 %)	71 ± 1.5	1.0 ± 0.9	70 ± 1.8	618 ± 3.
Fo + Cc + Ob (50 %)	76 ± 1.7	0.5 ± 0.7	0 ± 0	627 ± 3.
Fo + Pi + Ob (25 %)	51 ± 1.4	2.0 ± 0.8	87 ± 1.7	510 ± 2.
Fo + Lb + Ob (25 %)	48 ± 1.4	2.0 ± 1.0	82 ± 1.8	540 ± 1.
Fo + Cc + Ob (25 %)	46 ± 1.8	1.5 ± 0.7	0 ± 0	575 ± 2.

Values are the means (± SE) of 10 seedlings. Pi = *Paxillus involutus*, Lb = *Laccaria bicolor*, Cc = *Clitocybe clavipes*, Fm = *Fusarium moniliforme*, Fo = *F. oxysporum*, Ob = Oxine benzoate.

liforme and *F. oxysporum*; however, chemical protection was better than biological control alone. The synergistic effect involving integrated control using chemical and biological control may be more effective and longer lasting than the control achieved through either alone (OHR et al. 1973; ELAD et al. 1980; SCHROTH and HANCOCK 1981; CHAKRAVARTY et al. 1991). For successful integration of biological and chemical control of plant pathogens, the systems must be compatible. The potential of an integrated system is shown in our study, oxine benzoate at 170 µg/ml was not toxic to test fungi but was highly toxic to *F. moniliforme* and *F. oxysporum* at 1.7 µg/ml. Half and quarter rates of application of oxine benzoate plus *C. clavipes*, *L. bicolor*, or *P. involutus* increased seedling survival more than the recon-

treated with fungicide alone. Rigorous testing in the field is necessary to determine the interactions specific strains of beneficial fungi, pathogens, fungicides, and host plants and to determine the economic benefit of integrated biological and chemical control.

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Literature

- AGRIOS, G. N.: Plant Pathology. – Academic Press, San Diego, 1988.
- ARORA, D.: Mushrooms demystified. – Ten Speed Press, Berkeley, 1979.
- BAKER, K. F.: Evolving concepts of biological control of plant pathogens. – Ann. Rev. Phytopath. 26–85, 1987.
- BAKER, K. F., R. J. COOK: Biological control of plant pathogens. – Am. Phytopath. Soc., 1982.
- BLOOMBERG, W. J.: *Fusarium* root rot of Douglas-fir seedlings. – Phytopathology 63, 337–341, 1977.
- BOYLE, C. D., K. H. HELLENBRAND: Assessment of the effect of mycorrhizal fungi on drought tolerance of conifer seedlings. – Can. J. Bot. 69, 1764–1771, 1991.
- BOYLE, C. D., W. J. ROBERTSON, P. O. SALONIUS: Use of mycelial slurries of mycorrhizal fungi inoculum for commercial tree seedling nurseries. – Can. J. For. Res. 17, 1480–1486, 1987.
- CHAKRAVARTY, P., T. UNESTAM: Mycorrhizal fungi prevent disease in stressed pine seedlings. – Z. PflKrankh. PflSchutz 118, 335–340, 1987a.
- CHAKRAVARTY, P., T. UNESTAM: Differential influence of ectomycorrhizae on plant growth and disease resistance in *Pinus sylvestris* seedlings. – Z. PflKrankh. PflSchutz 120, 104–120, 1987b.
- CHAKRAVARTY, P., S. F. HWANG: Effect of an ectomycorrhizal fungus, *Laccaria laccata* on *Fusarium* damping-off in *Pinus banksiana* seedlings. – Eur. J. For. Path. 21, 97–106, 1991.
- CHAKRAVARTY, P., R. L. PETERSON, B. E. ELLIS: Integrated control of *Fusarium* damping-off in red pine seedlings. – Can. J. For. Res. 20, 1283–1288, 1990.
- CHAKRAVARTY, P., R. L. PETERSON, B. E. ELLIS: Interaction between the ectomycorrhizal fungi *Paxillus involutus* damping-off in *Pinus resinosa* seedlings. – Z. PflKrankh. PflSchutz 132, 207–211, 1991.
- DUCHESNE, L. C., R. L. PETERSON, B. E. ELLIS: The accumulation of plant-produced antimicrobial compounds in response to ectomycorrhizal fungi: a review. – Phytoprotection 68, 17–27, 1987.
- DUCHESNE, L. C., R. L. PETERSON, B. E. ELLIS: Interaction between the ectomycorrhizal fungi *Paxillus involutus* and *Pinus resinosa* induces resistance to *Fusarium oxysporum*. – Can. J. Bot. 66, 558–562, 1988a.
- DUCHESNE, L. C., R. L. PETERSON, B. E. ELLIS: Pine root exudate stimulates the synthesis of antifungal compounds by the ectomycorrhizal fungus *Paxillus involutus*. – New Phytol. 108, 470–476, 1988.
- DUCHESNE, L. C., R. L. PETERSON, B. E. ELLIS: The future of ectomycorrhizal fungi as biological control agents. – Phytoprotection 70, 51–58, 1989.
- ELAD, Y., I. CHET, J. KATAN: Physical, biological, chemical control integrated for soil-borne diseases in potatoes. – Phytopathology 70, 418–422, 1980.
- HIRATSUKA, Y., D. W. LANGOR, P. E. CRANE: A field guide to forest insects and diseases of the prairie provinces. – Can. For. Serv., North. For. Cent., Special Report 3, 1995.
- HWANG, S. F., P. CHAKRAVARTY, K. F. CHANG: The effect of two ectomycorrhizal fungi *Paxillia involutus* and *suillus tomentosus* and of *Bacillus subtilis* on *Fusarium* damping-off in Jack pine seedlings. – Phytoprotection 76, 57–66, 1995.
- MARX, D. H.: The influence of ectotrophic mycorrhizal fungi on the resistance of pathogenic infections. I. Antagonism of mycorrhizal fungi to pathogenic fungi and soil bacteria. – Phytopathology 59, 153–163, 1969.